

## Inflammation and Acute Venous Thrombosis

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### Abstract

Inflammation is known as a controlled response to an injury that protects against further injury and clears damaged tissue. However, uncontrolled inflammation can lead to a marked breakdown of the extracellular matrix as well as organ destruction. Stewart et al. demonstrated in a vein thrombosis model that leukocytes migrate into vein walls with an intact layer of endothelial cells, subsequently linking inflammation and thrombosis. Since then, multiple studies have reported on the interaction between inflammation and vein thrombosis. This article summarizes published material that assesses the relationship between inflammation and acute vein thrombosis. The article will focus on the role of endothelial cells, leukocytes, platelets, and microparticles in acute vein thrombosis and discuss the role of thrombin, P-selectin, platelet-activating factor, endothelin-1, and nitric oxide from the perspective of inflammation.

### Keywords

Inflammation, acute thrombosis, thrombin, selectin, leukocytes

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Venous thromboembolism (VTE) includes both deep vein thrombosis (DVT) and pulmonary embolism (PE). VTE occurs worldwide, and in all age groups and socioeconomic populations in North America and western Europe.<sup>1,2</sup> In a recent study by Heit et al., the estimated total annual cases of VTE occurring in the US exceeded 900,000, of which more than 250,000 were fatal.<sup>3</sup> Despite progress in medicine and biology, no changes in the incidence of VTE were noted during the 25-year cohort study.<sup>3</sup>

Elucidating the intrinsic mechanisms involved in thrombus formation is critical to understanding VTE pathophysiology. Stewart et al. demonstrated leukocyte migration into the vein wall with an intact layer of endothelial cells (ECs) during DVT,<sup>4</sup> linking inflammation and thrombosis. Furthermore, recent studies have reported on the interaction between inflammation and thrombosis.<sup>5–9</sup> This article summarizes published material to assess the relationship between inflammation and acute vein thrombosis. In particular, this article will focus on the role of ECs, leukocytes, platelets, microparticles, thrombin, P-selectin, platelet-activating factor, endothelin-1, and nitric oxide (NO) from the perspective of inflammation.

### Inflammation

Inflammation is a complex, highly regulated biological process that can be triggered by several stimuli, such as pathogens, noxious mechanical and chemical agents, and autoimmune responses.<sup>10</sup> In the clinical setting, inflammation is described as redness and swelling with heat and pain. Biologically, these signs and symptoms are explained by an

increasing microvascular caliber, enhanced vascular permeability, leukocyte recruitment, and the release of inflammatory mediators.<sup>10</sup>

Inflammation in physiologic states is a controlled response that protects against further injury and clears damaged tissue. In disease states such as sepsis, however, uncontrolled inflammation can lead to a marked breakdown of the extracellular matrix as well as organ destruction.<sup>10</sup> Inflammation can also be acute or chronic—two well-characterized states in which neutrophils and monocytes, respectively, play an important role. Similarly, DVT is a process with acute and chronic phases. In this setting, the main cells observed during acute DVT are the neutrophils. Monocytes are observed during the chronic phase. This parallel between inflammation and DVT is highly suggestive of a relationship.

### Inflammation and Vein Thrombosis

Using a vein thrombosis model in a dog, Stewart et al. showed that large numbers of white cells adhered to the vessel walls, passed through the endothelial intercellular junctions, and accumulated in pockets between the endothelium and basement membrane.<sup>4</sup> Since then, several studies have demonstrated the interaction between inflammation and thrombosis, noting the importance of ECs in this link.<sup>5–9</sup> The question of what activates ECs towards a procoagulant effect is fundamental in understanding the pathophysiology of VTE.

### Endothelial Cells

ECs play a pivotal role in venous thrombosis in healthy individuals. While arterial thrombosis requires EC disruption with collagen exposure (as

occurs in atherosclerotic plaque rupture), ECs are “intact and no collagen exposure is needed in order to generate thrombi during VTE.”<sup>11</sup> One of the main functions of ECs is to modulate the pro- and anticoagulant mechanisms. The anticoagulant properties of ECs include suppressing coagulation (platelet activation and adhesion) and inflammation (leukocyte activation).<sup>9</sup> Despite this, a procoagulant effect is observed during states of EC activation and disturbance, either physical (vascular trauma) or functional (sepsis).<sup>9</sup> Thus, under an adequate procoagulant stimulus, the balance between anti- and procoagulant mechanisms is altered toward procoagulant activity.<sup>1,12,13</sup>

### Thrombin

Thrombin (factor II) is a major procoagulant molecule that cleaves fibrinogen into fibrin, forming a fibrin mesh. Specifically, thrombin cleavage  $\alpha$  and  $\beta$  chain of fibrinogen releases fibrinopeptides A and B, respectively. This causes fibrin monomer polymerization that stabilizes the initial platelet plug. Thrombin also activates factor XIII, which contributes to thrombus amplification through the activation of platelet factors V and VIII.<sup>14</sup> In addition to this, thrombin itself has an inflammatory effect. Thrombin induces a rapid EC activation, increasing P-selectin expression and prostacyclin and platelet-activating factor secretion.<sup>15,16</sup> Thrombin may also activate transcription factor nuclear factor-kappa B (NF- $\kappa$ B) independently of the well-known pro-inflammatory cytokines tumor necrosis factor-alpha (TNF- $\alpha$ ) and interleukin-1 (IL-1).<sup>17,18</sup> This is critical in leukocyte activation and recruitment as it promotes EC secretion of monocyte chemoattractant protein-1 and IL-8.<sup>18,19</sup>

### P-selectin and Leukocytes

Selectins are cell-adhesion molecules that have critical roles in inflammation and thrombogenesis.<sup>5,9,20</sup> P-selectin is involved in leukocyte rolling and adhesion, which is an early inflammatory mechanism that facilitates leukocyte transmigration.<sup>4,20</sup> Animal studies utilizing rat and mouse thrombosis models have demonstrated the upregulation of P-selectin in the vein wall as early as six hours after thrombus induction.<sup>20</sup> These studies show that P-selectin is a common molecule that links inflammation and thrombosis *in vivo*.

The P-selectin receptor known as P-selectin glycoprotein ligand 1 (PSGL-1) is a glycoprotein expressed on the surface of leukocytes and platelets. It is key in the recruitment of leukocytes and platelets to inflamed tissue. The PSGL-1:P-selectin complex promotes rolling and adhesion of leukocytes and platelets, respectively, which ultimately results in increased vein wall cell infiltration.<sup>21-22</sup>

Chronologically, the initiation of inflammation and/or thrombosis processes occurs almost immediately after EC activation. P-selectin is stored inside the EC, within the Weibel-Palade bodies. Molecules of P-selectin are released upon EC activation, as reported by Wagner et al.<sup>23,24</sup> The exocytosis of Weibel-Palade bodies initiates a rapid translocation of P-selectin to the EC surface, resulting in its adhesiveness for leukocytes and platelets.

### Platelets

The role of platelets in arterial thrombosis is well known, but its action in venous thrombosis remains obscure. Platelets are anucleate circulating

blood particles derived from bone marrow megakaryocytes.<sup>2</sup> The megakaryocytes extend their cytoplasm into the bone marrow sinusoid and release a small portion of cytoplasm containing alpha and dense granules surrounded by a bi-lipid membrane into the blood circulation.

Under physiological conditions, circulating platelets are at rest (or non-activated), expressing PSGL-1.<sup>24,25</sup> As previously mentioned, thrombin promotes EC activation, resulting in P-selectin expression and platelet-activating factor secretion. EC P-selectin expression allows EC-platelet interaction (platelet PSGL-1:EC P-selectin), and platelets become activated by EC platelet-activating factor.<sup>21,24</sup> Ultimately, platelets excrete the contents of their granules, increasing platelet adhesiveness and potentiating platelet aggregation.<sup>25</sup> P-selectin is contained in platelet alpha granules. These granules are excreted with platelet activation, leading to P-selectin expression. These processes favor leukocyte-platelet cross-talk as a direct consequence of platelet activation.<sup>21,24</sup> It has been shown that platelets release a greater amount of P-selectin compared with ECs.<sup>24</sup> Thus, once platelets are attached to the vein wall, the concentration of available P-selectin increases dramatically, improving the leukocyte recruitment efficiency of the vein wall.<sup>24</sup> In a mouse model, Frenette et al. demonstrated that platelets roll on the vein wall (in a similar manner to leukocytes) after EC activation.<sup>21</sup>

### Microparticles

Circulating cell-derived microparticles are small vesicles (<1 $\mu$ m) consisting of plasma membrane surrounding a small amount of cytoplasm that contains cell-specific surface molecules that characterize their cell of origin.<sup>26,27</sup> Procoagulant microparticles derived from ECs, leukocytes, and platelets contribute to the coagulation and amplification of thrombosis.<sup>2,26,28</sup> Microparticles are present in the blood of healthy individuals but increase under specific circumstances, including DVT.<sup>2,29</sup> ECs, leukocytes, and platelets have a very well-structured plasma membrane characterized by a controlled transverse lipid distribution.<sup>30</sup> Specific areas of the plasma membrane have been identified as ‘lipid rafts,’<sup>31</sup> a formation of sphingolipids and cholesterol. It has been postulated that lipid rafts enhance signaling functions by providing platforms that physically concentrate receptors and downstream kinases involved in signaling pathways.<sup>32</sup> The membrane of microparticles is rich in lipid rafts.<sup>30</sup>

Moreover, lipid-raft-derived microparticles concentrate tissue factor.<sup>30</sup> Tissue factor and the expression of prothrombinase activity on the membrane and PSGL-1 are involved in the procoagulant activity of microparticles.<sup>9,33</sup> Therefore, the fusion of microparticles with activated platelets promotes thrombus formation in a tissue-factor-dependent manner.

During inflammation, the activation of ECs upregulates the expression of P-selectin on their surface, leading to the formation of EC P-selectin:PSGL-1-leukocyte complexes. These complexes stimulate the production of microparticles from leukocytes, particularly monocytes, along with platelets and ECs. Furthermore, the accumulation of leukocyte markers expressed on the surface of microparticles in the growing thrombi is mediated by the P-selectin:PSGL-1 complex,<sup>9</sup> resulting in a dramatic increase in microparticle concentration at the site of vein wall injury and inflammation.<sup>5</sup>

It has been suggested that microparticles participate in platelet activation. Recent reports have postulated that platelet activation may also result from the interaction of macrophage-1 antigen, found on leukocyte-derived microparticles, via microparticle:glycoprotein Ib (platelet) alpha polypeptide.<sup>34,35</sup>

In addition, a recent comparative proteomics analysis of microparticles showed that galectin-3 binding protein (Gal-3 BP) was significantly upregulated in DVT patients compared with negative controls.<sup>36</sup> Galectins are a family of carbohydrate-binding proteins that have a high affinity for the galactosides present on cell surfaces and extracellular glycoproteins.<sup>37</sup> Galectins are involved in multiple biological functions, including modulation of cell apoptosis, cell activation, and inflammation.<sup>37</sup> It has been suggested that galectins are involved in P-selectin expression.<sup>37</sup> Although the role of galectins and Gal-3 BP in thrombogenesis remains unclear, these observations suggest a new avenue linking inflammation and thrombosis.

## Interleukin-6

As previously mentioned, leukocyte diapedesis through an intact layer of endothelial cells has been described in early stages of venous thrombosis (VT), linking acute inflammation and acute thrombosis.<sup>38-42</sup> This event further incites the inflammatory response and alters the composition of local cytokines. Increased plasma levels of interleukin-6 (IL-6), a major inflammatory cytokine, have been demonstrated during VT<sup>43</sup> and post-thrombotic syndrome (PTS)<sup>44</sup>. While these studies suggest that IL-6 may be used as a marker for VT, its role as an active cytokine in the development of PTS has been recently established.<sup>45</sup> Using the mouse inferior vena cava (IVC) ligation model to generate vein thrombosis and the neutralization of IL-6, with an antibody anti-IL6, we studied the role of IL-6 in VT. We found that IL-6 had an acute and a chronic effect in VT. During the acute phase of thrombosis, CCL-2 was significantly decreased, two days after thrombosis in the group where circulating IL-6 was blocked. This was demonstrated at the gene expression level as well as at the protein level.

Moreover, the decrease in CCL-2 caused a reduction in the number of monocytes recruited into the injured area, at an intermediate time-point (six days after VT) indicating a dynamic and linked process. At the chronic time-point (14 days after surgery), a statistically significant decrease in fibrosis was observed in the group treated with anti-IL6 versus control. This was demonstrated using several methods, including Masson's stain quantification of fibrosis. In summary, we have shown a relationship between IL-6, CCL-2, monocyte recruitment, and fibrosis for the first time in the context of VT, exhibiting a potential pathway that started early on in VT and continued to have an impact on chronic VT as well.<sup>45</sup> This is an important step in understanding the link between inflammation and thrombosis in the context of VT, and indeed a potential therapeutic target for PTS.

## Platelet-activating Factor

Platelet-activating factor, also known as acetyl-glycerol-ether-phosphorylcholine (AGEPC), is a molecule secreted by ECs, macrophages, mast cells, and leukocytes.<sup>46</sup> The thrombotic and pro-inflammatory effects of platelet-activating factor have been suggested in several studies. In an animal study using baboons,

thrombocytopenia and neutropenia were observed secondary to the intravenous infusion of platelet-activating factor, suggesting both thrombotic and pro-inflammatory effects.<sup>47</sup> In addition, platelet-activating factor was shown to cause aggregation and accumulation of platelets and subsequent changes in local blood flow at the sites of experimental thrombosis *in vivo*.<sup>48</sup>

One of the main roles of platelet-activating factor during inflammation is to activate the leukocytes adhered to the vessel wall, in a paracrine fashion, via adhesion molecules expressed by ECs.<sup>46,49</sup> Activation of leukocytes induced by platelet-activating factor leads to priming for enhanced inflammatory responses, polarization and directional migration, degranulation and oxygen radical generation.

## Endothelin-1

Endothelin-1 (ET-1) is a 21 amino acid peptide produced by a variety of cells including endothelial and smooth-muscle cells.<sup>50</sup> ET-1 receptors include ET<sub>A</sub> and ET<sub>B</sub>.<sup>50-51</sup> ET<sub>A</sub> is expressed in inflammatory cells.<sup>52</sup> Specific locations for ET<sub>B</sub> receptors have been suggested: in vein ECs for ET<sub>B1</sub> and vein vascular smooth-muscle cells.<sup>50</sup> Once ET-1 binds to its receptor (Gq-proteins) on the vascular smooth muscle it induces an increase in inositol 1,4,5-triphosphate levels, with subsequent calcium release and muscle contraction.<sup>50</sup> Specifically, it has been shown that EC dysfunction and inflammation contribute to an overproduction of ET-1 in humans.<sup>52</sup> The exact role of ET-1 in VTE, however, remains unclear. Future studies are needed to elucidate the role of ET-1 in venous thrombosis.

## Prostacyclin and Nitric Oxide

Finally, prostacyclin and NO are secreted by ECs.<sup>53</sup> These molecules contribute synergistically to vessel homeostasis by reducing the tone and growth of vascular smooth-muscle cells, platelet aggregation, and leukocyte adhesion to the endothelium, thus decreasing the vessel's susceptibility to form thrombi.<sup>2,53</sup> Interestingly, Osanai et al. demonstrated that vessel homeostasis might be maintained through an increase in prostacyclin production in vascular ECs when NO synthesis is impaired.<sup>54</sup> Endothelial NO synthase function has been widely studied in arterial ECs, but there is also evidence suggesting that decreased NO production may play a role in the development of venous disease.<sup>55-57</sup>

## Summary

This article has reviewed the current evidence of inflammatory mechanisms in vein thrombosis. The Conrad Jobst Vascular Laboratory at the University of Michigan is currently studying other inflammatory molecules, such as galectins and inflammatory cytokines. As previously mentioned, Gal-3 BP was upregulated in human microparticles obtained from patients undergoing DVT.<sup>37</sup>

Acute VTE is a complex and dynamic process. ECs, platelets, and leukocytes are the main circulatory elements involved in vein thrombosis. Inflammation has been shown to be closely involved in vein thrombus formation. Inflammatory cytokines and thrombin are thought to orchestrate this early phase of DVT. Further understanding of the intrinsic mechanism involved in DVT may help to develop new preventive and/or therapeutic modalities directed to these potential targets. ■

1. Kitchens CS, Alving BM, Kessler CM, *Consultative Hemostasis and Thrombosis*, Philadelphia: WB Saunders Co., 2002.
2. Heith JA, Thrombophilia: clinical and laboratory assessment and management. In: Kitchens CS, Alving BM, Kessler CM (eds), *Consultative Hemostasis and Thrombosis*, Philadelphia: WB Saunders Co., 2002:213–44.
3. Silverstein MD, Heit JA, Mohr DN, et al., Trends in the incidence of deep vein thrombosis and pulmonary embolism: a 25-year population-based study, *Arch Intern Med*, 1998;158(6):585–93.
4. Stewart GJ, Ritchie WG, Lynch PR, Venous endothelial damage produced by massive sticking and emigration of leukocytes, *Am J Pathol*, 1974;74(3):507–32.
5. Myers DD, Wakefield TW, Inflammation-dependent thrombosis, *Front Biosci*, 2005;10:2750–7.
6. Collins T, Read MA, Neish AS, et al., Transcriptional regulation of endothelial cell adhesion molecules: NF-kappa B and cytokine-inducible enhancers, *FASEB J*, 1995;9(10):899–909.
7. Chen W, Esselman WJ, Jump DB, et al., Anti-inflammatory effect of docosahexaenoic acid on cytokine-induced adhesion molecule expression in human retinal vascular endothelial cells, *Invest Ophthalmol Vis Sci*, 2005;46(11):4342–7.
8. Murase T, Kume N, Hase T, et al., Gallates inhibit cytokine-induced nuclear translocation of NF-kappaB and expression of leukocyte adhesion molecules in vascular endothelial cells, *Arterioscler Thromb Vasc Biol*, 1999;19(6):1412–20.
9. Wakefield TW, Myers DD, Henke PK, Mechanisms of venous thrombosis and resolution, *Arterioscler Thromb Vasc Biol*, 2008;28(3):387–91.
10. Firestein G, Mechanisms of inflammation and tissue repair. In: Goldman L, Ausiello D (eds), *Cecil Textbook of Medicine*, Philadelphia: Saunders, 2004:227–33.
11. Mackman N, Tilley RE, Key NS, Role of the extrinsic pathway of blood coagulation in hemostasis and thrombosis, *Arterioscler Thromb Vasc Biol*, 2007;27(8):1687–93.
12. Gamble JR, Harlan JM, Klebanoff SJ, et al., Stimulation of the adherence of neutrophils to umbilical vein endothelium by human recombinant tumor necrosis factor, *Proc Natl Acad Sci U S A*, 1985;82(24):8667–71.
13. Schleimer RP, Rutledge BK, Cultured human vascular endothelial cells acquire adhesiveness for neutrophils after stimulation with interleukin 1, endotoxin, and tumor-promoting phorbol diesters, *J Immunol*, 1986;136(2):649–54.
14. Stubbs MT, Bode W, A player of many parts: the spotlight falls on thrombin's structure, *Thromb Res*, 1993;69(1):1–58.
15. Lorant DE, Patel KD, McIntyre TM, et al., Coexpression of GMP-140 and PAF by endothelium stimulated by histamine or thrombin: a juxtacrine system for adhesion and activation of neutrophils, *J Cell Biol*, 1991;115(1):223–34.
16. Weksler BB, Ley CW, Jaffe EA, Stimulation of endothelial cell prostacyclin production by thrombin, trypsin, and the ionophore A 23187, *J Clin Invest*, 1978;62(5):923–30.
17. Rahman A, Anwar KN, True AL, et al., Thrombin-induced p65 homodimer binding to downstream NF-kappa B site of the promoter mediates endothelial ICAM-1 expression and neutrophil adhesion, *J Immunol*, 1999;162(9):5466–76.
18. Kaplanski G, Fabrigoule M, Boulay V, et al., Thrombin induces endothelial type II activation in vitro: IL-1 and TNF-alpha-independent IL-8 secretion and E-selectin expression, *J Immunol*, 1997;158(11):5435–41.
19. Colotta F, Sciacca FL, Sironi M, et al., Expression of monocyte chemoattractant protein-1 by monocytes and endothelial cells exposed to thrombin, *Am J Pathol*, 1994;144(5):975–85.
20. Myers D Jr, Farris D, Hawley A, et al., Selectins influence thrombosis in a mouse model of experimental deep venous thrombosis, *J Surg Res*, 2002;108(2):212–21.
21. Frenette PS, Johnson RC, Hynes RO, et al., Platelets roll on stimulated endothelium in vivo: an interaction mediated by endothelial P-selectin, *Proc Natl Acad Sci U S A*, 1995;92(16):7450–4.
22. Myers DD Jr, Henke PK, Bedard PW, et al., Treatment with an oral small molecule inhibitor of P selectin (PSI-697) decreases vein wall injury in a rat stenosis model of venous thrombosis, *J Vasc Surg*, 2006;44(3):625–32.
23. Bonfanti R, Furie BC, Furie B, et al., PADGEM (GMP140) is a component of Weibel-Palade bodies of human endothelial cells, *Blood*, 1989;73(5):1109–12.
24. Wagner DD, Frenette PS, The vessel wall and its interactions, *Blood*, 2008;111(11):5271–81.
25. Jung SM, Platelet collagen receptors. In: Tanaka K, Davie EW (eds), *Recent Advances in Thrombosis and Hemostasis*, Springer, 2008:231–42.
26. Blann A, Shantsila E, Shantsila A, Microparticles and arterial disease, *Semin Thromb Hemost*, 2009;35(5):488–96.
27. Ahn ER, Lander G, Jy W, et al., Differences of soluble CD40L in sera and plasma: implications on CD40L assay as a marker of thrombotic risk, *Thromb Res*, 2004;114(2):143–8.
28. Nomura S, Ozaki Y, Ikeda Y, Function and role of microparticles in various clinical settings, *Thromb Res*, 2008;123(1):8–23.
29. Enjeti AK, Lincz LF, Seldon M, Microparticles in health and disease, *Semin Thromb Hemost*, 2008;34(7):683–91.
30. Del Conde II, Shrimpton CN, Thiagarajan P, et al., Tissue-factor-bearing microvesicles arise from lipid rafts and fuse with activated platelets to initiate coagulation, *Blood*, 2005;106(5):1604–11.
31. Simons K, Ikonen E, Functional rafts in cell membranes, *Nature*, 1997;387(6633):569–72.
32. Pierce SK, Lipid rafts and B-cell activation, *Nat Rev Immunol*, 2002;2(2):96–105.
33. Satta N, Toti F, Feugeas O, et al., Monocyte vesiculation is a possible mechanism for dissemination of membrane-associated procoagulant activities and adhesion molecules after stimulation by lipopolysaccharide, *J Immunol*, 1994;153(7):3245–55.
34. Pluskota E, Woody NM, Szpak D, et al., Expression, activation, and function of integrin alphaMbeta2 (Mac-1) on neutrophil-derived microparticles, *Blood*, 2008;112(6):2327–35.
35. Andrews RK, Berndt MC, Microparticles facilitate neutrophil/platelet crosstalk, *Blood*, 2008;112(6):2174–5.
36. Ramacciotti E, Hawley AE, Wroblewski SK, et al., Proteomics of microparticles after deep venous thrombosis, *Thromb Res*, 2010;125(6):e269–e74.
37. Diaz JA, Ramacciotti E, Wakefield TW, Do galectins play a role in venous thrombosis? a review, *Thromb Res*, 2009;125(5):373–6.
38. Roumen-Klappe EM, Janssen MC, Van Rossum J, et al., Inflammation in deep vein thrombosis and the development of post-thrombotic syndrome: a prospective study, *J Thromb Haemost*, 2009;7(4):582–7.
39. Wakefield TW, Linn MJ, Henke PK, et al., Neovascularization during venous thrombosis organization: a preliminary study, *J Vasc Surg*, 1999;30(5):885–92.
40. Wakefield TW, Strieter RM, Schaub R et al., Venous thrombosis prophylaxis by inflammatory inhibition without anticoagulation therapy, *J Vasc Surg*, 2000;31(2):309–24.
41. Henke PK, Varma MR, Deatrick KB, et al., Neutrophils modulate post-thrombotic vein wall remodeling but not thrombus neovascularization, *Thromb Haemost*, 2006;95(2):272–81.
42. Varma MR, Varga AJ, Knipp BS, et al., Neutropenia impairs venous thrombosis resolution in the rat, *J Vasc Surg*, 2003;38(5):1090–8.
43. Roumen-Klappe EM, et al., Inflammatory response in the acute phase of deep vein thrombosis, *J Vasc Surg*, 2002;35(4):701–6.
44. Shbaklo H, Holcroft CA, Kahn SR, Levels of inflammatory markers and the development of the post-thrombotic syndrome, *Thromb Haemost*, 2009;101(3):505–12.
45. Wojcik BM, Wroblewski SK, Hawley AE, et al., Interleukin-6: A Potential Target for Post-thrombotic Syndrome, *Ann Vasc Surg*, 2011;25(2):229–39.
46. Tjoelker LW, Stafforini DM, Platelet-activating factor acetylhydrolases in health and disease, *Biochim Biophys Acta*, 2000;1488(1–2):102–23.
47. McManus LM, Kinckard RN, Fitzpatrick FA, et al., Acetyl glyceryl ether phosphorylcholine. Intravascular alterations following intravenous infusion into the baboon, *Lab Invest*, 1981;45(4):303–7.
48. Golino P, Ambrosio G, Ragni M, et al., Short-term and long-term role of platelet activating factor as a mediator of *in vivo* platelet aggregation, *Circulation*, 1993;88(3):1205–14.
49. Zimmerman GA, Lorant DE, McIntyre TM, et al., Juxtacrine intercellular signaling: another way to do it, *Am J Respir Cell Mol Biol*, 1993;9(6):573–7.
50. Watts SW, Endothelin receptors: what's new and what do we need to know?, *Am J Physiol Regul Integr Comp Physiol*, 2009;298(2):R254–60.
51. Ram CV, Possible therapeutic role of endothelin antagonists in cardiovascular disease, *Am J Ther*, 2003;10(6):396–400.
52. Mencarelli M, Pecorelli A, Carbotti P, et al., Endothelin receptor A expression in human inflammatory cells, *Regul Pept*, 2009;158(1–3):1–5.
53. Sessa WC, eNOS at a glance, *J Cell Sci*, 2004;117(Pt 12):2427–9.
54. Osanai T, Akutsu N, Fujita N, et al., Cross talk between prostacyclin and nitric oxide under shear in smooth muscle cell: role in monocyte adhesion, *Am J Physiol Heart Circ Physiol*, 2001;281(1):H177–82.
55. Higman DJ, Strachan AM, Butterly L, et al., Smoking impairs the activity of endothelial nitric oxide synthase in saphenous vein, *Arterioscler Thromb Vasc Biol*, 1996;16(4):546–52.
56. Higman DJ, Greenhalgh RM, Powell JT, Smoking impairs endothelium-dependent relaxation of saphenous vein, *Br J Surg*, 1993;80(10):1242–5.
57. Broeders MA, Tangelder GJ, Slaaf DW, et al., Endogenous nitric oxide protects against thromboembolism in venules but not in arterioles, *Arterioscler Thromb Vasc Biol*, 1998;18(1):139–45.