

Dynamics of Human Papillomavirus and Cervical Cancer Screening

Channa E Schmeink,¹ Leon FAG Massuger,¹ Willem JG Melchers²
and Ruud LM Bekkers¹

1. Department of Obstetrics and Gynaecology; 2. Department of Medical Microbiology, Radboud University Nijmegen Medical Centre

Abstract

Primary screening based on detection of human papillomavirus (HPV) has proved to be more sensitive than cytology for the detection of high-grade cervical intraepithelial neoplasia (CIN). Self-sampling for specimen collection may also improve the participation rate, especially in the non-responder group. However, HPV is highly prevalent and therefore HPV detection has a lower specificity in cervical cancer screening than cytology. In addition to the clinically validated HPV test, HPV dynamics should be taken into account. It is important to identify women with a chronic productive infection likely to cause, or to already have caused, high-grade CIN or cervical carcinoma, and to limit overtreatment of women with a transient infection. Furthermore, the introduction of the HPV vaccine is likely to lower the incidence of CIN and cervical carcinoma, which will lower the positive predictive value of cervical cancer screening. This potential impact needs to be taken into account when planning for future screening guidelines.

Keywords

Human papillomavirus (HPV), cervical cancer screening, transmission, dynamics, latency

Disclosure: The authors have no conflicts of interest to declare.

Received: 26 June 2011 **Accepted:** 8 August 2011 **Citation:** *European Oncology & Haematology*, 2011;7(4):243–6 DOI: 10.17925/EOH.2011.07.04.243

Correspondence: Channa E Schmeink, Radboud University Nijmegen Medical Centre, Department of Obstetrics and Gynaecology (internal mail 791), PO Box 9101, 6500 HB Nijmegen, The Netherlands. E: c.schmeink@obgyn.umcn.nl

The introduction of human papillomavirus (HPV) testing into cervical screening programmes seems to be inevitable. However, HPV is a highly prevalent virus that is mainly transient. Therefore, it is important to understand HPV dynamics and to identify women at high risk of cervical intraepithelial neoplasia (CIN) or cervical carcinoma to avoid overtreatment. Persistent HPV infection is necessary for the development of high-grade CIN or cervical carcinoma.^{1,2} Fortunately, in most individuals the immune response eventually leads to clearance of HPV, or to its maintenance in a latent or asymptomatic state in epithelial basal cells. It remains to be elucidated which proportion of HPV infection clears or is maintained in a latent state. Latency implies that the virus may remain within basal epithelial cells, either arrested or very slowly replicating, but barely detectable by current DNA technology. Short-term fluctuations in HPV and fluctuations in HPV throughout the menstrual cycle^{3,4} raise the following questions:

- When an HPV type is detected, when does a woman have a productive (replicating) infection?
- When is the detected HPV type infectious? (i.e., when is there a high risk of transmission to a sexual partner?)
- When is a newly detected HPV type truly a new infection and when does it represent a re-infection or reactivation?

To answer these questions, we must consider the role of the immune system and the role of the viral load.

Productive Human Papillomavirus Infection

The presence of HPV may indicate a productive (replicating) infection or not. It is important to realise that an infection is more than the

detection of the pathogen. Infection is invasion by, and multiplication of, pathogenic microorganisms in bodily tissue, which may produce tissue injury and progress to disease through a variety of cellular or toxic mechanisms.

When studying the HPV life-cycle, initial infection requires access of infectious particles to cells in the basal layer through microabrasions. After cell division, one daughter cell migrates and undergoes differentiation. This induces the productive phase, in which expression of E6 and E7 deregulates cell-cycle control, allowing viral genome amplification. On reaching the uppermost layer of the epithelium, the virions are shed.^{5,6} At this point, depending on the amount of virions shed (i.e., viral load), the virus may be detected and may be transmitted to others. In women who do not successfully resolve their chronic productive HPV infection, CIN may develop and progress to CIN3 and, eventually, to cervical carcinoma (i.e. clinically relevant infection).^{7,8}

Some studies have shown that an increased viral load is associated with an increased risk of clinically relevant cervical lesions.^{9,10} However, this potential relationship between viral load and disease is not shown for every HPV type. Studies mainly showed that HPV type 16 viral load increases with increasing disease severity.^{10–12} Snijders et al. showed that viral load was also significantly higher for HPV types 18, 31 and 33 in scrapes of women with \geq CIN2 compared with those of women with normal cytology.⁹ On the other hand, longitudinal studies showed that baseline viral load did not relate to outcome of CIN2/3 and that only changes in viral load correlated with risk of CIN 2/3 development.^{13,14} Furthermore, there exists a wide

range in viral DNA load levels and a substantial overlap between women with and without CIN3.^{10,15} Setting a clinically significant viral load cut-off value that is predictive for disease is therefore difficult and viral load is not the preferred tool in clinical practice.

Transmission of Human Papillomavirus

The immune system is important in the natural control of the spread of HPV-associated disease. The immune system may clear the infection or control the virus by keeping it at a low copy number.

Transmission dynamics are dependent on both pathogen and host factors and are defined by three components:

- transmissibility from an infected to an uninfected partner upon exposure;
- the likelihood of exposure between infected and uninfected persons; and
- the time for which a person is infectious.¹⁶

When studying genital HPV prevalence, the main factors associated with HPV prevalence are related to sexual behaviour.¹⁷ In a simulation model based on likelihood of sexual exposures between infected and uninfected persons and the duration of the infection, the transmissibility of genital HPV from an infected to an uninfected partner upon exposure was estimated to be a median of 40 % per sexual act. The probability of male-to-female transmission would reach virtually 100 % within only 11 acts of sexual intercourse. This estimated high rate of transmission of HPV implies that the potential protective effect of condoms would disappear over multiple acts of sexual intercourse.¹⁶ However, condom use does partially prevent HPV transmission and therefore reduces the spread of HPV in a sexually active population. It even may promote HPV clearance and regression of CIN and penile lesions.^{18,19}

Studies analysing HPV transmission in couples showed that HPV concordance is higher in couples with cervical or penile lesions present and that a higher genital viral load increases the transmission rate.²⁰⁻²³ These results on HPV transmission underline the importance of the sexual partner in the viral dynamics of high-risk HPV infection of the genital tract.

Human Papillomavirus – When Is It Newly Acquired, a Re-infection or a Reactivation?

Short-term fluctuations of HPV in an individual suggest that some detected HPV types might be newly acquired whereas others might have been acquired in the past and remained latent, below detection level for a certain period, and then been reactivated.

Reactivation of latent HPV infections was observed in HIV-infected women; however, few reactivation events were identified in HIV-uninfected women.²⁴ The most important factor consistently associated with reactivation in HIV-infected women is a CD4 count less than 200 mm³.²⁴ This suggests that functional immune systems keep HPV infections in a sub-clinical state and that they may be reactivated by immunosuppressive conditions.

Unfortunately, we do not know how frequently latency occurs among immune-competent individuals, how long it may last, what causes reactivation into a detectable state and what fraction of cancer arises after a period of HPV latency. Because of the apparently low rate

of reactivation, large studies would be needed to adequately address reactivation in an immune-competent group.²⁴ Studying HPV reactivation in cervical infection is also complicated by the inability to distinguish reactivation of an existing infection from re-infection with the same HPV type through sexual contact with an infected partner. Therefore, knowledge of previous infection, sexual behaviour within the HPV testing interval and testing the sexual partner are required to exclude the possibility of re-infection. Furthermore, it is unclear whether clearance of carcinogenic HPV infections may result in type-specific immunity and whether immunity needs boosting over time.^{20,25,26} Moreover, only about 50–60 % of women with carcinogenic HPV infection develop detectable serum antibodies.^{20,21,25,26}

Some studies report a relation between viral load and seroconversion of immunoglobulin G (IgG),^{12-19,24,25} whereas others have identified persistent HPV infection to be a significant factor.^{21,22} A high viral load may provide an acute and rapid immune response, whereas a persistent infection maintains a slow gradual boosting effect.²⁵ Another factor potentially related to seroconversion of IgG is oral contraceptive use.^{22,25} Hormones may induce transcription of the integrated viral oncogenes and influence the mucosal immunity in the genital tract.²³ The influence of the endogenous and exogenous hormones on the mucosal immune response most likely explains the differences in HPV detection within and between the oral contraceptive cycle and the natural menstrual cycle.³

This variety in data on HPV seroprevalence demonstrates that the host immune response against HPV is only partially understood and that HPV serology is a poor marker of current infections or related lesions.

Human Papillomavirus DNA Detection in Cervical Cancer Screening

To date, HPV DNA testing has been used in most cervical screening programmes as a triage test for women who have abnormal cytology. However, HPV DNA-based primary screening has proved to be more sensitive than cytology in detecting high-grade CIN, but with a lower specificity.^{27,28} Additionally, HPV DNA-based primary screening may also improve the participation rate, especially when self-sampling is used for specimen collection, thereby increasing the overall effectiveness of the screening programme.^{29,30} Self-sampling has been proved to have a sufficient sensitivity to screen women otherwise not screened, but further research is necessary before wide implementation in a national screening programme is possible.

To improve the specificity of HPV DNA-based primary screening, HPV dynamics should be taken into account.² HPV dynamics can indicate whether a detected high-risk HPV type can be considered as:

- a newly acquired infection, or re-infection, or reactivation, that could be a transient infection or potentially persisting and causing CIN within several years;
- an already persistent infection likely to cause/have caused a CIN or cervical carcinoma (i.e., a chronic productive infection); or
- an accidental pick-up of a latent infection.

HPV persistence as a clinical marker may be of value to identify women who are at high risk of cervical cancer. Repeat (genotype-specific) HPV DNA testing at 12 months may therefore be a valuable tool to identify women at increased risk of CIN and cervical cancer.³¹ HPV genotyping might be used as an additional tool to decide whether to treat or not,

because some types are more likely to persist than others.³¹ Especially for women under 30 years of age, it will be important for clinicians to be conservative and wait for evidence of a persistent (chronic productive) high-risk HPV infection rather than act on the first positive test for high-risk HPV, because these women are more likely to have a transient infection and the lesion (if present) may regress.³¹⁻³⁴

Novel biomarkers – immunologic, genetic or viral factors – may be useful for triage in HPV DNA detection-based primary screening to support decision-making on treatment or a ‘wait and see’ policy.^{35,36} Currently developed biomarkers are E6 and E7 HPV messenger RNA transcripts, methylation of several genes, markers for viral integration and of chromosomal instability, and markers of increased cell proliferation such as Ki-67 and p16.³⁷⁻³⁹

The high negative predictive value of the HPV DNA test for developing CIN3 means that very few relevant lesions are missed.^{27,40} However, studies have shown that, in natural HPV dynamics, a sequence of positive high-risk HPV samples may be interrupted by a single high-risk HPV-negative sample.^{41,42} Within these fluctuations, we may assume either that the viral load is below detection level (depending on the replication rate of the virus, latency or the sensitivity of the HPV DNA test used) or that there may be a sampling error. When a non-productive high-risk HPV infection is ‘missed’, there are most probably no clinical consequences. However, when an active high-risk HPV infection is missed owing to the low sensitivity of an HPV test or sampling error, this leads to inadequate follow-up and treatment. Thus, to safely extend this cervical screening interval in case of a negative test, the sensitivity of the HPV DNA detection test used in primary cervical cancer screening should be sufficiently high to detect CIN3 or a chronic productive high-risk HPV infection likely to produce CIN3 within the screening interval.

Influence of Human Papillomavirus (HPV) Vaccination on HPV Dynamics

As an increasing proportion of the population is vaccinated, the prevalence of cervical abnormalities will decrease. However, given the lag in time between HPV infection and the development of cervical cancer, (universal) vaccination is likely to have only a minimal impact on CIN and cervical cancer rates until 10–20 years from now.⁴³ Additionally, vaccination against HPV cannot provide 100 % protection against cervical carcinoma and its precursor lesions and little is known about the potential long-term benefit of (cross-)type immune response. Therefore, vaccinated women still need to be followed and cervical cancer screening programmes must continue.

The potential impact of HPV vaccination on the epidemiology of HPV and the rate of abnormal cytology depends on the following variables:

- vaccination coverage;
- the duration of vaccine-induced immune protection;
- the target high-risk HPV types of the vaccine;
- the rate of cross-protection; and
- whether HPV type replacement takes place.

In 2009, the UK and Australia used school-based vaccination programmes and achieved a high three-dose completion rate of approximately 80 and 70 %, respectively.^{44,45} In the Netherlands, however, only 52 % of the girls in the target age group received the HPV vaccine. When vaccination coverage is less than 90 %, herd

immunity obtained by only vaccinating women may be insufficient to eradicate the targeted HPV types.⁴⁶ When considering vaccinating boys to increase herd immunity, the potential gain of a further reduction of cervical cancer must also be carefully weighed against the extra costs. It appears that, when female programmes obtain high (over 75 %) coverage, the vaccination of males provides only a small additional benefit and is not cost-effective.⁴⁷

In the future, long-term follow-up studies should determine the true efficacy and duration of vaccine-induced immune protection of both vaccines. However, to date there have been no cases of infection or cytological lesions associated with HPV16/18 in 7.3 years of follow-up after vaccination with the bivalent vaccine (HPV-16/18 AS04-adjuvanted: Cervarix®; GlaxoSmithKline, North Carolina, US).⁴⁸ Vaccination with the quadrivalent vaccine (HPV- 6/11/16/18: Gardasil®; Merck & Co., New Jersey, US) had a high prophylactic efficacy against low-grade cervical and vulvovaginal neoplasia and condylomata associated with the vaccine types through 42 months of follow-up; vaccine efficacy against CIN1 was 96 %.⁴⁹

The percentage of all cervical cancers attributable to HPV 16/18 in Europe has been estimated at 71 %.⁵⁰ The percentage of prevented cervical cancer is potentially higher as both vaccines have reported cross-protection against other high-risk HPV types. As summarised by Szarewski,⁵¹ the efficacy provided by the bivalent vaccine against CIN2+ lesions was 100 % for HPV-31 or -45, 66.1 % for non-HPV-16 α -9 species and 77.3 % for non-HPV-18 α -7 species;⁵² the efficacy provided by the quadrivalent vaccine against CIN2+ lesions was 58.7 % for HPV-31 or -45, 35.4 % for non-HPV-16 α -9 species and 47.0 % for non-HPV-18 α -7 species.⁵³ Although these results are encouraging, the duration of cross-protection is unknown.

Currently, the efficacy of a broad-spectrum vaccine (Merck) against nine HPV types, including seven oncogenic HPV types, is being studied in a randomised phase III trial. When a polyvalent vaccine against nine HPV types is implemented, the prevalence of the majority of cervical cancer-associated HPV types will be drastically decreased and the discussion on cross-protection will change.

There is a possibility that the distribution of HPV types may gradually change in vaccinated populations to fill the vacated ecologic niches after the elimination of HPV types 16 and 18.^{54,55} Type replacement is a viral population dynamics phenomenon and is defined as elimination of some types causing an increase in incidence of other types. This effect can occur only if two conditions apply:

- there exists partial competition among different types during natural infection; and
- the vaccine does not afford cross-protection against types affected by this natural competition.⁵⁵

Future epidemiologic studies are needed to show whether type replacement occurs.

Challenges to Primary Cervical Cancer Screening After Vaccination

The potential impact of HPV vaccination needs to be taken into account when planning for future screening guidelines. It is estimated that there will be a 50–60 % reduction in colposcopy referrals owing to prevention of HPV 16 and 18 lesions. This will lower the positive predictive value of

any screening test for cervical cancer.⁵⁶ However, HPV testing is automated and therefore more objective, and thus is less likely to be influenced by this effect than cytology.⁵⁷ Owing to the high costs of vaccination, it is important to restrict the costs of cervical screening programmes and implement an approach based on vaccine implementation.⁵⁸ However, later and less frequent screening will probably become an acceptable worldwide policy only if vaccination uptake is high across all social and economic strata.⁵⁹ To help develop new guidelines for cervical cancer screening algorithms in the

post-vaccine era, future epidemiologic studies should monitor HPV prevalence, type replacement, the level of protection against HPV 16 and 18, the level of cross-protection, the duration of protection and the population coverage of the vaccination.

In the future, provided there is high vaccination coverage with a polyvalent vaccine or a high level of cross-protection, a long duration of protection or a validated booster vaccination schedule, screening will likely be superfluous. ■

- Nobbenhuis MA, Walboomers JM, Helmerhorst TJ, et al., Relation of human papillomavirus status to cervical lesions and consequences for cervical-cancer screening: a prospective study, *Lancet*, 1999;354:20-5.
- Bekkers RL, Massuger LF, Bulten J, Melchers WJ, Epidemiological and clinical aspects of human papillomavirus detection in the prevention of cervical cancer, *Rev Med Virol*, 2004;14:95-105.
- Schmeink CE, Massuger LF, Lenselink CH, et al., Effect of the menstrual cycle and hormonal contraceptives on human papillomavirus detection in young, unscreened women, *Obstet Gynecol*, 2010;116:67-75.
- Van Ham MA, Melchers WJ, Hanselaar AG, et al., Fluctuations in prevalence of cervical human papillomavirus in women frequently sampled during a single menstrual cycle, *Br J Cancer*, 2002;87:373-6.
- Doorbar J, The papillomavirus life cycle, *J Clin Virol*, 2005;32(Suppl. 1):S7-15.
- Moody CA, Laimins LA, Human papillomavirus oncoproteins: pathways to transformation, *Nat Rev Cancer*, 2010;10:550-60.
- Trottier H, Franco EL, The epidemiology of genital human papillomavirus infection, *Vaccine*, 2006;24(Suppl. 1):S1-15.
- Kjaer SK, Frederiksen K, Munk C, Iftner T, Long-term absolute risk of cervical intraepithelial neoplasia grade 3 or worse following human papillomavirus infection: role of persistence, *J Natl Cancer Inst*, 2010;102:1478-88.
- Snijders PJ, Hogewoning CJ, Hesselink AT, et al., Determination of viral load thresholds in cervical scrapings to rule out CIN 3 in HPV 16, 18, 31 and 33-positive women with normal cytology, *Int J Cancer*, 2006;119:1102-7.
- Swan DC, Tucker RA, Tortolero-Luna G, et al., Human papillomavirus (HPV) DNA copy number is dependent on grade of cervical disease and HPV type, *J Clin Microbiol*, 1999;37:1030-4.
- Gravitt PE, Burk RD, Lorincz A, et al., A comparison between real-time polymerase chain reaction and hybrid capture 2 for human papillomavirus DNA quantitation, *Cancer Epidemiol Biomarkers Prev*, 2003;12:477-84.
- Tabora N, Ferrera A, Bakkers JM, et al., High HPV 16 viral load is associated with increased cervical dysplasia in Honduran women, *Am J Trop Med Hyg*, 2008;78:843-6.
- Clavel C, Masure M, Lever M, et al., Human papillomavirus detection by the hybrid capture II assay: a reliable test to select women with normal cervical smears at risk for developing cervical lesions, *Diagn Mol Pathol*, 2000;9:145-50.
- Crum CP, Beach KJ, Hedley ML, et al., Dynamics of human papillomavirus infection between biopsy and excision of cervical intraepithelial neoplasia: results from the ZYC101a protocol, *J Infect Dis*, 2004;189:1348-54.
- Snijders PJ, Steenbergen RD, Heideman DA, Meijer CJ, HPV-mediated cervical carcinogenesis: concepts and clinical implications, *J Pathol*, 2006;208:152-64.
- Burchell AN, Richardson H, Mahmud SM, et al., Modeling the sexual transmissibility of human papillomavirus infection using stochastic computer simulation and empirical data from a cohort study of young women in Montreal, Canada, *Am J Epidemiol*, 2006;163:534-43.
- Lenselink CH, Melchers WJ, Quint WG, et al., Sexual behaviour and HPV infections in 18 to 29 year old women in the pre-vaccine era in the Netherlands, *PLoS ONE*, 2008;3:e3743.
- Bleeker MC, Berkhof J, Hogewoning CJ, et al., HPV type concordance in sexual couples determines the effect of condoms on regression of flat penile lesions, *Br J Cancer*, 2005;92:1388-92.
- Winer RL, Hughes JP, Feng Q, et al., Condom use and the risk of genital human papillomavirus infection in young women, *N Engl J Med*, 2006;354:2645-54.
- Frazer IH, Interaction of human papillomaviruses with the host immune system: a well evolved relationship, *Virology*, 2009;384:410-4.
- Carter JJ, Koutsky LA, Hughes JP, et al., Comparison of human papillomavirus types 16, 18, and 6 capsid antibody responses following incident infection, *J Infect Dis*, 2000;181:1911-9.
- Wideroff L, Schiffman MH, Hoover R, et al., Epidemiologic determinants of seroreactivity to human papillomavirus (HPV) type 16 virus-like particles in cervical HPV-16 DNA-positive and-negative women, *J Infect Dis*, 1996;174:937-43.
- Brabin L, Interactions of the female hormonal environment, susceptibility to viral infections, and disease progression, *AIDS Patient Care STDS*, 2002;16:211-21.
- Theiller RN, Farr SL, Karon JM, et al., High-risk human papillomavirus reactivation in human immunodeficiency virus-infected women: risk factors for cervical viral shedding, *Obstet Gynecol*, 2010;115:1150-8.
- Ho GY, Studentsov YY, Bierman R, Burk RD, Natural history of human papillomavirus type 16 virus-like particle antibodies in young women, *Cancer Epidemiol Biomarkers Prev*, 2004;13:110-6.
- Viscidi RP, Schiffman M, Hildesheim A, et al., Seroreactivity to human papillomavirus (HPV) types 16, 18, or 31 and risk of subsequent HPV infection: results from a population-based study in Costa Rica, *Cancer Epidemiol Biomarkers Prev*, 2004;13:324-7.
- Dillner J, Rebolj M, Birembaut P, et al., Long term predictive values of cytology and human papillomavirus testing in cervical cancer screening: joint European cohort study, *BMJ*, 2008;337:a1754.
- Cuzick J, Sasieni P, Davies P, et al., A systematic review of the role of human papilloma virus (HPV) testing within a cervical screening programme: summary and conclusions, *Br J Cancer*, 2000;83:561-5.
- Bekkers RL, Meijer CJ, Massuger LF, et al., Effects of HPV detection in population-based screening programmes for cervical cancer; a Dutch moment, *Gynecol Oncol*, 2006;100:451-4.
- Gok M, Heideman DA, van Kemenade FJ, et al., HPV testing on self collected cervicovaginal lavage specimens as screening method for women who do not attend cervical screening: cohort study, *BMJ*, 2010;340:c1040.
- Castle PE, Rodriguez AC, Burk RD, et al., Short term persistence of human papillomavirus and risk of cervical precancer and cancer: population based cohort study, *BMJ*, 2009;339:b2569.
- Moscicki AB, Conservative management of adolescents with abnormal cytology and histology, *J Natl Compr Canc Netw*, 2008;6:101-6.
- Ostor AG, Natural history of cervical intraepithelial neoplasia: a critical review, *Int J Gynecol Pathol*, 1993;12:186-92.
- Moscicki AB, Ma Y, Wibbelsman C, et al., Rate of and risks for regression of cervical intraepithelial neoplasia 2 in adolescents and young women, *Obstet Gynecol*, 2010;116:1373-80.
- Schiffman M, Wentzensen N, Wacholder S, et al., Human papillomavirus testing in the prevention of cervical cancer, *J Natl Cancer Inst*, 2011;103:368-83.
- van HD, Bulten J, Shirango H, et al., Biological behavior of CIN lesions is predictable by multiple parameter logistic regression models, *Carcinogenesis*, 2008;29:840-5.
- Cuschieri K, Wentzensen N, Human papillomavirus mRNA and p16 detection as biomarkers for the improved diagnosis of cervical neoplasia, *Cancer Epidemiol Biomarkers Prev*, 2008;17:2536-45.
- Wentzensen N, Sherman ME, Schiffman M, Wang SS, Utility of methylation markers in cervical cancer early detection: appraisal of the state-of-the-science, *Gynecol Oncol*, 2009;112:293-9.
- Boulet GA, Horvath CA, Berghmans S, Bogers J, Human papillomavirus in cervical cancer screening: important role as biomarker, *Cancer Epidemiol Biomarkers Prev*, 2008;17:810-7.
- Ronco G, Giorgi-Rossi P, Carozzi F, et al., Efficacy of human papillomavirus testing for the detection of invasive cervical cancers and cervical intraepithelial neoplasia: a randomised controlled trial, *Lancet Oncol*, 2010;11:249-57.
- Insinga RP, Perez G, Wheeler CM, et al., Incidence, duration, and reappearance of type-specific cervical human papillomavirus infections in young women, *Cancer Epidemiol Biomarkers Prev*, 2010;19:1585-94.
- Moscicki AB, Ma Y, Jonte J, et al., The role of sexual behavior and human papillomavirus persistence in predicting repeated infections with new human papillomavirus types, *Cancer Epidemiol Biomarkers Prev*, 2010;19:2055-65.
- Cuzick J, Long-term cervical cancer prevention strategies across the globe, *Gynecol Oncol*, 2010;117(2 Suppl.):S11-4.
- Brotherton JM, Deeks SL, Campbell-Lloyd S, et al., Interim estimates of human papillomavirus vaccination coverage in the school-based program in Australia, *Commun Dis Intell*, 2008;32:457-61.
- Cuzick J, Castanon A, Sasieni P, Predicted impact of vaccination against human papillomavirus 16/18 on cancer incidence and cervical abnormalities in women aged 20-29 in the UK, *Br J Cancer*, 2010;102:933-9.
- Berkhof J, Bogaards JA, Vaccination against human papillomavirus types 16 and 18: the impact on cervical cancer, *Future Oncol*, 2010;6:1817-21.
- Marra F, Cloutier K, Oteng B, et al., Effectiveness and cost effectiveness of human papillomavirus vaccine: a systematic review, *Pharmacoeconomics*, 2009;27:127-47.
- De CN, Teixeira J, Roteli-Martins CM, et al., Sustained efficacy and immunogenicity of the HPV-16/18 AS04-adjuvanted vaccine up to 7.3 years in young adult women, *Vaccine*, 2010;28:6247-55.
- Dillner J, Kjaer SK, Wheeler CM, et al., Four year efficacy of prophylactic human papillomavirus quadrivalent vaccine against low grade cervical, vulvar, and vaginal intraepithelial neoplasia and anogenital warts: randomised controlled trial, *BMJ*, 2010;341:c3493.
- Clifford G, Franceschi S, Diaz M, et al., Chapter 3: HPV type-distribution in women with and without cervical neoplastic diseases, *Vaccine*, 2006;24(Suppl. 3):S3-26-S3/34.
- Szarewski A, HPV vaccine: Cervarix, *Expert Opin Biol Ther*, 2010;10:477-87.
- Paavonen J, Naud P, Salmeron J, et al., Efficacy of human papillomavirus (HPV)-16/18 AS04-adjuvanted vaccine against cervical infection and precancer caused by oncogenic HPV types (PATRICIA): final analysis of a double-blind, randomised study in young women, *Lancet*, 2009;374:301-14.
- Brown DR, Kjaer SK, Sigurdsson K, et al., The impact of quadrivalent human papillomavirus (HPV) types 6, 11, 16, and 18) L1 virus-like particle vaccine on infection and disease due to oncogenic nonvaccine HPV types in generally HPV-naive women aged 16-26 years, *J Infect Dis*, 2009;199:926-35.
- Franco EL, Trottier H, Reassessing the epidemiology of human papillomavirus infection: back to basics, *Sex Transm Dis*, 2008;35:283-5.
- Stanley M, Lowy DR, Frazer I, Chapter 12: Prophylactic HPV vaccines: underlying mechanisms, *Vaccine*, 2006; 24(Suppl. 3):S3-106-S3/113.
- van Hamont D., Bekkers RL, Massuger LF, Melchers WJ, Detection, management, and follow-up of pre-malignant cervical lesions and the role for human papillomavirus, *Rev Med Virol*, 2008;18:117-32.
- Cuzick J, Arbyn M, Sankaranarayanan R, et al., Overview of human papillomavirus-based and other novel options for cervical cancer screening in developed and developing countries, *Vaccine*, 2008;26(Suppl. 10):K29-K41.
- Basu P, Chowdhury D, Cervical cancer screening & HPV vaccination: a comprehensive approach to cervical cancer control, *Indian J Med Res*, 2009;130:241-6.
- Massad LS, Einstein M, Myers E, et al., The impact of human papillomavirus vaccination on cervical cancer prevention efforts, *Gynecol Oncol*, 2009;114:360-4.