

## Translating Promising Biomarkers into Oncology Clinical Practice

Tracy G Lively, PhD,<sup>1</sup> Michael C Sachs, PhD<sup>2</sup> and Barbara A Conley, MD<sup>3</sup>

1. Deputy Associate Director, Cancer Diagnosis Program, Division of Cancer Treatment and Diagnosis, National Cancer Institute, National Institutes of Health, Department of Health and Human Services; 2. Mathematical Statistician, Biometric Research Branch Program, Division of Cancer Treatment and Diagnosis, National Cancer Institute, National Institutes of Health, Department of Health and Human Services; 3. Associate Director, Cancer Diagnosis Program, Division of Cancer Treatment and Diagnosis, National Cancer Institute, National Institutes of Health, Department of Health and Human Services, Bethesda, US

### Abstract

There is a clear need for biomarker assays that can assist in the choice of the best treatment for individual cancer patients. Many manuscripts can be found in the literature that describe promising correlations of molecular features with cancer outcomes, but few of these ever translate to assays that come into common clinical use. Some reasons for this lack of development are the failure to define a clear and compelling intended use for the biomarker assay, as well as the paucity of necessary and careful validation studies: the analytical and clinical validation of a given assay, optimization for clinical use, and demonstration of the clinical utility of the assay. These steps often require resources beyond those of the original laboratory that defined the correlation of the biomarker result to clinical outcome. This manuscript details some of the matters that are faced by assay developers, as well as some solutions for improved assay development and validation.

### Keywords

Tumor biomarker, clinical assay development, clinical utility, assay validation, precision medicine

**Disclosure:** Tracy G Lively, PhD, Michael C Sachs, PhD, and Barbara A Conley, MD, have no conflicts of interest to declare. No funding was received in the publication of this article.

**Open Access:** This article is published under the Creative Commons Attribution Noncommercial License, which permits any noncommercial use, distribution, adaptation, and reproduction provided the original author(s) and source are given appropriate credit.

**Received:** January 28, 2015 **Accepted:** March 16, 2015 **Citation:** *Oncology & Hematology Review*, 2015;11(1):14–8 DOI: 10.17925/OHR.2015.11.01.14

**Correspondence:** Barbara A Conley, MD, Associate Director, Cancer Diagnosis Program Division of Cancer Treatment and Diagnosis, NCI, NIH, DHHS, 9609 Medical Center Drive, Room 4W426, MSC9609, Bethesda, MD 20892, US. E: conleyba@mail.nih.gov

Although the literature is replete with biomarker studies that show promise for predicting risk for cancer, response to treatment, or prognosis after diagnosis, the number of biomarker-based tests or assays that are useful in the practice of oncology remains distressingly small.<sup>1–3</sup> A biomarker can be used to predict a patient's prognosis, which is their expected clinical outcome. A biomarker that provides accurate predictions of prognosis, regardless of treatment, is referred to as prognostic. A predictive biomarker is one that accurately predicts disease outcomes with the application of specific interventions or treatments. Predictive biomarkers are therefore useful for the selection among two or more treatment options. There are many clinical decisions that could plausibly be informed by a reliable biomarker or assay so the rationale for development of biomarkers remains great.

For example, for estimating cancer risk, we can now predict that certain heritable mutations confer increased risk for cancer, such as *BRCA 1* or *2*. The clinical decision that the assay informs is how to mitigate that risk, either by surgical removal of the organ most at risk, increased invasive or imaging surveillance, or long-term chemopreventive medical treatment. The inheritance of a *BRCA 1* or *2* mutation conveys a risk for 18–57 % for the development of breast or ovarian cancers.<sup>4</sup> For better preventive

planning, biomarkers that could define the risk for cancer more accurately for the individual patient (not just the population of patients with a particular mutation) would also be very useful. Most cancers are not heritable, and for these, it would be desirable to have biomarkers to inform if that cancer in a particular patient will be aggressive, or indolent (prognostic biomarker). Other clinical scenarios in which informative predictive biomarkers would be welcome include the choice of the most efficacious course of adjuvant therapy after surgery for curative intent in an individual patient, the best choice of systemic or radiation therapy in patients with advanced, incurable cancers, and the risk for severe toxicity of a given treatment in that patient.

Some cancers now require the measurement of several predictive biomarkers for treatment planning.<sup>5</sup> For example, breast cancer requires the assessment of estrogen and progesterone receptor and *HER2* amplification. Lung cancer requires the assessment of activating *EGFR* mutations and *ALK* translocations. With the advent of next-generation sequencing, we are also faced with how to make the best decision when the data to support action in the presence of a particular molecular abnormality or set of abnormalities are sparse. Such assays may be used routinely in the near future for “personalized” or “precision” treatment of cancer patients—tailoring treatment to each patient's unique molecular profiles.<sup>6</sup>

The association of a molecular abnormality or feature with a particular clinical outcome is the beginning of the process for many predictive and prognostic tests. However, further development of that insight to a reliable and accurate clinical assay is complex. Most tests or assays do not survive the process for various reasons. Significant technical knowledge and resources are required for success—such knowledge and resources must be specifically sought, and are not always present in the team that initially proposes the promising association. Effective planning must occur to develop an assay properly for use in a clinical test; and missteps can cause significant confusion that persists for years. Worst of all, if a test that is not properly developed is used in clinical care, inaccurate treatment and potential harm to patients could result. If diagnostics are essential to choosing the appropriate treatment (sometimes termed companion diagnostics), a similar degree of attention should be given to their development as is given to the development of cancer drugs. Definitions of terms used in this review are given in the Text Box at the end of this article.

### Assay Development

When developing an assay, it is essential to identify a specific intended use. An assay will likely not be equally useful for discerning cancer aggressiveness and choosing the best treatment, nor are assays that measure a particular biomarker equally useful across different cancers (e.g. breast cancer and colon cancer), since the amount of biomarker present, as well as the cutpoint for positivity may vary across tumors. The development of an assay and its intended use should address a clinical need, and a particular clinical decision. At the beginning of development, once a correlation of the assay result with a clinical outcome has been discerned, the developer must consider whether more information about the association is actually needed before committing considerable resources to continue assay development and validation. How will the assay help a clinical decision? What decision will the assay guide? Is this an important decision, not addressed by current tests or clinical judgment? The answers to these questions will help guide the clinical development of the assay, including the design of clinical trials, the patient populations, the type of specimen that will be used, the assay platform, etc.

Additional practical factors will need to be considered. Does the assay measure a prognostic or a predictive marker, or both? How large is the population of patients for which that assay will be used, and what is the prevalence of the analyte(s) that the assay detects? If the assay is for a predictive marker or a companion diagnostic, what would the effect of the targeted treatment be on patients who are “negative” for the marker? What is the turnaround time necessary for clinical use? What type of specimen is needed for the assay and what handling/preservation/storage procedures will be fit for the purpose of the assay as well as for feasibility of clinical use? What is the cost of the assay?

If there is already a “research assay,” will the research assay and platform be suitable for use in the clinical situation? For example, an immunohistochemistry test that detected a relationship between an analyte and survival when it was performed on a tissue microarray will not be suitable for the clinic if results were normalized using the median value obtained on that array. If the analysis was carried out on many samples at once, will the assay perform on samples that arrive one at a time from the clinic? If the assay requires a tissue sample that must be frozen or fresh, or obtained and placed in preservative within a short time of obtaining

the sample, is this feasible for use in the clinic? Will the sample degrade over time? If so, specific pre-analytic techniques must be developed and validated.<sup>7</sup> If an antibody is used to detect the analyte, does that antibody perform as well if a different batch of the antibody is used? If the current research assay will not translate to clinical workflow easily, it is reasonable to assess other platforms and conditions for better suitability.

### Analytical Validation

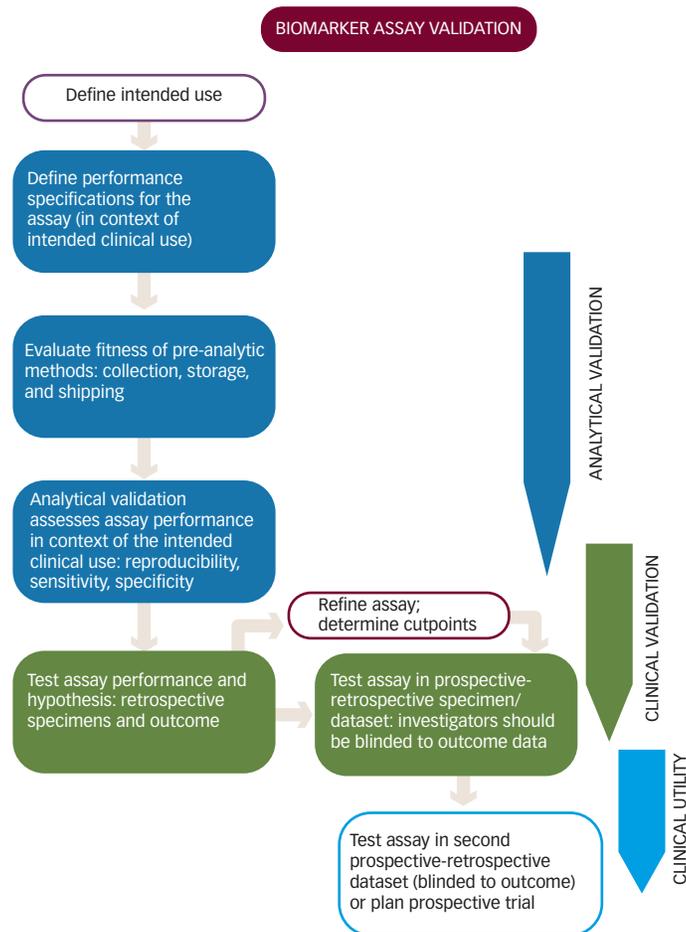
Analytical validity is the assessment of how well the assay detects the desired analyte in the population in which it is intended to be used. Prior to use in the clinic or to use in a clinical validation study (see below) an assay must be analytically validated. Analytical validation involves assessment of accuracy, precision, lower limit of detection, linearity, robustness (interfering substances, robustness to various sample handling procedures), range of performance, and reproducibility across instruments, by different technicians and by different laboratories if more than one laboratory will be performing an assay in a clinical trial.<sup>7-9</sup> The analytical validation plan will assess how well the assay performs in the context of its intended clinical use. The analytical validation will also assess whether the assay can meet performance metrics that have been defined to be necessary for clinical use of that assay. For example, if an assay to detect minimal residual disease needs to detect one malignant cell in one million cells, the assay validation must assess how well it will do this. The assessment includes how well the assay detects an analyte when it is present (sensitivity), and how often the assay result is negative when in fact the analyte is not present (specificity). The analytical validation plan must also ascertain how often the assay is “positive” when the analyte is not present (false positive) and how often the assay is negative when in fact the analyte is present (false negative). The developer must decide what the reasonable rates (limits) of false positive and false negative results should be, given how the assay is intended to be used. The assay must then be developed to “fit” these demands. Once the analytical validation is complete, assay procedures, reagents, software, etc. should be “locked down”—this means there should be no further changes to the method after the assay validation.

### Clinical Validation

Clinical validity refers to how well the assay result, the biomarker, relates to the clinical outcome of interest, such as frequency of relapse, response to therapy, or survival. Here consideration must be given to how the assay will be used to be sure the assessment of its clinical validity is carried out in the appropriate patient population. Assessment of clinical validity relates to the both analytical performance and to the intended clinical use. The assessment of clinical validity, particularly, can be an iterative process (see *Figure 1*). If the analytical performance of the assay has been designed well, the analytical validation will not likely have to be repeated unless there are changes or upgrades to the assay or pre-analytic requirements. Of course, if the measurement of the analyte or signature no longer correlates at all with clinical outcome when used in a new dataset, the assay itself may need to be redeveloped.

Prospective-retrospective clinical validations refer to the type of study that assesses the performance of the assay on clinical samples from patients who participated in a (completed) clinical trial, and for whom the outcome of the treatment in the clinical trial is known. Prospective-retrospective assessments are sometimes ideal for clinical validation, if they can be performed rigorously in a sample from a well-defined patient population.<sup>10</sup>

**Figure 1: Schematic of Steps that May be Involved in Evaluating Analytical Validity, Clinical Validity, and Clinical Utility of a New Biomarker Assay**



Assessment of clinical validity for an assay that measures multiple analytes, such as “omics” assays, can become complex, and the developer must understand certain pitfalls in the development of such predictors.<sup>11–13</sup> Likewise, continuous-valued biomarkers that require a “cutpoint” that defines “positive” and “negative” may require several sets of clinical data to refine and lock down the cutpoint before use in the clinic. A common pitfall in biomarker studies is optimizing the cutpoint and then evaluating its performance using the same data. This causes the performance estimates to be optimistically biased.<sup>14</sup> This type of bias, called resubstitution bias, can be avoided by carefully separating the data used to optimize the biomarker from that used to assess its clinical utility. This can be accomplished by dividing the data and using one set to optimize the biomarker, and the other set to assess its clinical utility. Adaptive study designs have been proposed in which the biomarker cutpoint is optimized and evaluated in the same trial.<sup>14</sup> Resampling methods such as cross-validation are commonly used to prevent resubstitution bias and provide valid estimates of the biomarker performance.

## Clinical Utility

Clinical utility is the determination of whether use of the biomarker assay for decision-making leads to improved clinical results for patients. Does the biomarker add to techniques already in use? Will the use of the biomarker

result in a different, beneficial treatment for the patient? If a “biomarker-positive” assay result predicts a group of patients that benefit more from a treatment than the biomarker-negative group, but the biomarker-negative group still derives as much or more benefit from the treatment as they would with any other treatment, there would be no reason to perform the assay for such a treatment decision. On the other hand, if a biomarker predicts benefit of a treatment for one or more subgroups of patients, while defining a group that will not benefit or may even have a worse outcome, that biomarker would be very useful for guiding treatment decisions (see *Figure 2*).

Prospective-retrospective studies may also inform about clinical utility. If a rigorous protocol for the prospective-retrospective analysis is prespecified and followed, then a high level of evidence supporting the clinical utility of the biomarker can be generated.<sup>10</sup> This strategy allowed, for example, the validation of the clinical utility of detecting KRAS mutation in patients with metastatic colorectal cancer. Prospective-retrospective analysis of tissues from patients with metastatic colorectal cancer who participated in several randomized studies of the efficacy of EGFR monoclonal antibodies (cetuximab or panitumumab) showed that such patients were not likely to benefit from treatment with EGFR monoclonal antibodies if their tumor had a KRAS mutation.<sup>15</sup>

Another example in which prospective-retrospective studies were used to address clinical utility was the development of the breast cancer classifier OncotypeDx™. This assay uses reverse transcription polymerase chain reaction (RT-PCR) to assess the expression of 16 cancer-related genes and five reference genes in paraffin embedded early stage, node negative, estrogen receptor-positive tumors to predict the likelihood of distant recurrence.<sup>16,17</sup> A locked down assay and prespecified study plan were used to evaluate tamoxifen-treated patients from a completed trial of adjuvant tamoxifen versus placebo in node negative, estrogen receptor-positive breast cancer patients. The assay was able to separate patients into low, intermediate, and high risk for distant recurrence-free survival at 10 years. A subsequent prospective-retrospective study evaluated whether high-risk patients, as evaluated by the assay, derived benefit from adjuvant chemotherapy. In this study, samples that had been banked from node negative, estrogen receptor-positive breast cancer patients who participated in a randomized adjuvant clinical trial to evaluate the worth of adding chemotherapy to tamoxifen treatment were used. The study showed that the patients classified as high risk by the assay derived benefit from adding chemotherapy to tamoxifen.<sup>18</sup> Finally, a prospective study (Clinicaltrials.gov Identifier NCT00310180) was conducted to evaluate whether patients classified as intermediate risk by the assay had better distant disease-free survival by adding chemotherapy to hormone therapy in the adjuvant setting. This trial has completed accrual and final results are pending.<sup>19</sup>

Other methods of assessing clinical utility involve the use of “convenience” samples and data. These data may be from series of patients who have not been on clinical trials, but may have been treated similarly and have outcome information, such as patients in a database of an oncology practice or group of practices. Here, one must be cautious of bias that may be present because treatment decisions not made randomly, but potentially based on some unrecorded patient characteristic, or because

of provider bias about treatments. If the samples and data have been collected over several decades, both the disease characteristics and treatments may have changed over time.

In any retrospective trial, tissue or other samples may not have been collected and stored in an ideal way for the biomarker assay, and so assessment of their fitness for the assay must be performed.

In the planning of a retrospective study, careful consideration should be given to the study power and sample size as precious archived specimens should not be wasted on a retrospective study that is not likely to evaluate the clinical validity of an assay.

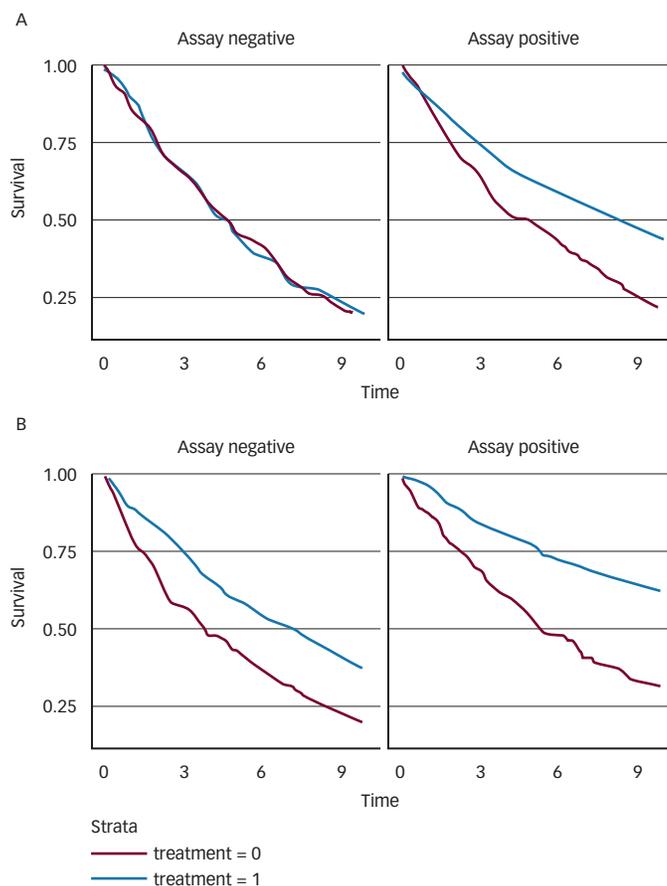
Finally, a prospective assessment (prospective clinical trial or study) could be done to assess clinical utility, which does allow specification of tissue or sample collection and storage, but the timeline may be longer than if retrospective samples/data can be identified. A variety of study designs exist to prospectively evaluate a potentially prognostic or predictive biomarker for clinical utility.<sup>20-27</sup> The effective integration of biomarker test development into clinical trials of investigational therapeutic agents requires both planning and multidisciplinary collaboration.<sup>22-23</sup> The type of study used to evaluate a biomarker, such as an enrichment or all-comers design, depends on the level and quality of prior evidence, both empirical and theoretical, supporting its utility. The study should have a clear and comprehensive prespecified protocol, including specimen-handling procedures, assay procedures, and statistical analysis plans. Investigators should prespecify a statistic or set of statistics that address the intended clinical use of the biomarker. In the design, careful consideration should be given to the study power and sample size. A prospective study in which patients may be undergoing painful biopsies or unnecessary treatment should not be done unless there is a high probability of definitively answering the scientific question.

## Other Considerations

Some necessary steps in test development may be in tension with other goals in the design of clinical trials. For example, a biomarker may be proposed as a predictive marker. If a well-characterized assay with an established cutpoint is available to identify the patient population of interest, it can be used even in early-phase trials to enrich enrollment for likely responders and limit the numbers of patients exposed to an investigational agent.<sup>25</sup> But if the assay itself is under development, then establishing that cutpoint and the relevant clinical sensitivity and specificity of the test requires assessing the treatment outcome in at least some marker-negative as well as marker-positive cases.<sup>24,25</sup> Even for a truly predictive biomarker, care must be taken not to place excessive confidence in a research grade assay for restricting enrollment to a trial, because of the effects that an imperfect assay can have on development efficiency. False positive cases will decrease statistical power by decreasing the effect size; false negative test results will slow accrual by screening out patients who should have been entered. In some cases it may be necessary to decide whether to prioritize sensitivity (minimizing false negatives) or specificity (minimizing false positives) for a clinical trial.

As another example, there can be tension between the goal of proceeding as quickly as possible from one phase to the next in the evaluation of an especially promising agent, and the need to allow time for the necessary

**Figure 2: Two Hypothetical Predictive Assays**



Panel A shows an assay that is predictive and clinically useful for determining whether to treat or not. In this hypothetical randomized controlled trial, the assay negative population does not benefit from treatment while the assay positive population does benefit. Panel B shows an assay that is predictive but is not clinically useful. The treatment is beneficial in both assay groups, but to a different degree (hazard ratio = 0.56 for negative; = 0.41 for positive). Despite the fact that there is a treatment by assay interaction, the assay is not clinically useful because regardless of the assay result, the correct clinically decision would be to treat.

steps in assay development to take place. Two-stage trial designs have been proposed to streamline drug and biomarker co-development, designs in which a biomarker is tested but not used to restrict enrollment in stage one and then, only if needed and if a well-validated assay has been developed, becomes an eligibility criterion in stage two.<sup>21</sup> These designs are most applicable to the transition between a randomized phase II and a phase III trial, that is, for later stage trials, and it may not be realistic to expect a fully “seamless” transition from stage one to stage two within the same trial. Such an expectation presupposes that the assay has been fully developed and validated to the point where it is fit for use as an eligibility criterion, and has an Investigational Device Exemption from the US Food and Drug Administration, if necessary,<sup>22</sup> even though it might never be needed for stage two. Time may also be required for assessment of interlaboratory comparability if multiple testing laboratories will be involved.

Other aspects of biomarker test development can be accelerated if efforts are initiated in early phase trials specifically for this purpose. Information about biomarker prevalence, assay performance in the tissue of interest, and specimen-handling requirements can be acquired even

before phase I from studies on nontrial specimens or tumor xenografts. As biomarkers are selected for inclusion as correlative laboratory studies in a trial, consideration can be given to how readily a research-grade assay, if developed further, could be incorporated into clinical practice. These considerations include not only the prospect of integrating the assay into the standard clinical workflow, but also the level of interest from commercial partners and anticipated regulatory requirements.

## Conclusions

In summary, bringing predictive and prognostic biomarkers in to clinical use requires a thoughtful development plan beginning with the careful consideration of how the assay will inform clinical decision-making, through assay development, and ending with demonstration of clinical utility. Careful planning and execution can assure that the assay is analytically validated, meaning that it measures the biomarker with accuracy and reproducibility. Once analytically validated, it is of interest to show that assay results correlate with the outcome of interest; that it has clinical validity. Throughout the whole process, investigators should keep in mind the clinical need that they intend to address. To show clinical utility, feasibility in the clinic must be considered, including properties of the sample that will be used and the platform, and that the assay use could bring a benefit to the patient. ■

## Definitions

**Analytical validity:** The extent to which the test detects the analyte of interest, in the clinical context (i.e. stage of disease, specimen type)

**Assay:** A process by which a molecular feature or set of features is measured in a biospecimen

**Clinical utility:** The extent to which the use of the test improves the clinical outcome of interest

**Clinical validity:** The strength of the association between the analyte of interest and the clinical outcome of interest

**Molecular biomarker:** A molecular feature that “is objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes, or pharmacologic responses to a therapeutic intervention”<sup>28</sup>

**Predictive biomarker:** a biomarker that predicts response/outcome to a particular treatment; e.g. HER2 amplification and trastuzumab

**Prognostic biomarker:** A biomarker that predicts outcome regardless of treatment, e.g. OncotypeDx for estrogen receptor-positive early stage breast cancer

- Ludwig JA, Weinstein JN, Biomarkers in cancer staging, prognosis and treatment selection, *Nature Reviews Cancer*, 2005;5:845–56.
- Khelif SN, Doroshow JH, Hait WN for the AACR-FDA-NCI-Biomarkers Collaborative, AACR-NCI-FDA Cancer Biomarkers Collaborative Consensus Report: Advancing use of biomarkers in cancer drug development, *Clin Cancer Res*, 2010;16:3299–318.
- Hayes DF, Biomarker validation and testing, *Mol Oncol*, 2014 [Epub ahead of print].
- Chen S, Parmagiani G, Meta-analysis of BRCA 1 and BRCA2 penetrance, *J Clin Oncol*, 2007;25:1329–33.
- Buettner R, Wolf J, Thomas RK, Lessons learned from lung cancer genomics: emerging concept of individualized diagnostics and treatment, *J Clin Oncol*, 2013;31:1858–65.
- Cardiello F, Arnold D, Casali PG, et al., Delivering precision medicine in oncology today and in future—the promise and challenges of personalised cancer medicine: a position paper by the European Society for Medical Oncology (ESMO), *Ann Oncol*, 2014;25:1673–78.
- Hewitt SM, Badve SS, True LD, Impact of preanalytic factors on the design and application of integral biomarkers for directing patient therapy, *Clin Cancer Res*, 2012;18:1524–30.
- Lee JW, Devanarayan V, Barrett YC, et al., Fit for purpose method development and validation for successful biomarker measurement, *Pharm Res*, 2006;23:312–28.
- Cummings J, Raynaud F, Jones L, et al., Fit for purpose biomarker method validation for application in clinical trials of anticancer drugs, *Br J Cancer*, 2010;103:1313–7.
- Simon RM, Paik S, Hayes DF, Use of archived specimens in evaluation of prognostic and predictive biomarkers, *J Natl Cancer Inst*, 2009;101:1446–52.
- McShane LM, Cavenagh MM, Lively TG, et al., Criteria for the use of omics-based predictors in clinical trials, *Nature*, 2013;502:317–20.
- Simon R, Radmacher MD, Dobbin K, McShane LM, Pitfalls in the use of DNA microarray data for diagnostic and prognostic classification, *J Natl Cancer Inst*, 2010;95:14–8.
- Subramanian J, Simon R, Gene expression based prognostic signatures in lung cancer: Ready for clinical use?, *J Natl Cancer Inst*, 2010;102:464–74.
- Freidlin B, Jiang W, Simon R, The cross-validated adaptive signature design, *Clin Cancer Res*, 2010;16:691–8.
- Normanno N, Tejpar S, Morgillo F, et al., Implications for KRAS status and EGFR targeted therapies in metastatic CRC, *Nat Rev Clin Oncol*, 2009;6:519–27.
- Paik S, Shak S, Tang G, et al., A multigene assay to predict recurrence of tamoxifen treated, node negative breast cancer, *New Engl J Med*, 2004;351:2817–26.
- Cronin M, Sangli C, Liu M-L, et al., Analytical validation of the oncotype dX genomic diagnostic test for recurrence prognosis and therapeutic response prediction in node-negative, estrogen receptor-positive breast cancer, *Clin Chem*, 2007;53:1084–91.
- Paik S, Tang G, Shak S, et al., Gene expression and benefit of chemotherapy in women with node negative, estrogen receptor-positive breast cancer, *J Clin Oncol*, 2006;24:3726–34.
- Zujewski JA, Kamin L, Trial assessing individualized options for treatment for breast cancer: the TailorX trial, *Future Oncology*, 2008;4:603–10.
- Freidlin B, McShane LM, Korn EL, Randomized clinical trials with biomarkers: design issues, *J Natl Cancer Inst*, 2010;102:152–60.
- Freidlin B, McShane LM, Polley MY C, Korn EL, Randomized phase II trial designs with biomarkers, *J Clin Oncol*, 2012;30:3304–9.
- Schilsky RL, Doroshow JH, LeBlanc M, et al., Development and use of integral assays in clinical trials, *Clin Cancer Res*, 2012;18:1540–6.
- Polley MY-C, Freidlin B, Korn EL, et al., Statistical and practical considerations for clinical evaluation of predictive biomarkers, *J Natl Cancer Inst*, 2013;105:1677–83.
- Hoering A, LeBlanc M, Crowley JJ, Randomized phase III clinical trial designs for targeted agents, *Clin Cancer Res*, 2008;14:4358–67.
- Dancey JE, Dobbin KK, Groshen S et al. Guidelines for the development and incorporation of biomarker studies in early clinical trials of novel agents, *Clin Cancer Res*, 2010;16:1745–55.
- Atkinson AJ Jr, Colburn WA, DeGruttola VG, et al., Biomarkers and surrogate endpoints: preferred definitions and conceptual framework, *Clin Pharmacol Ther*, 2007;69:89–95.
- Teutsch SM, Bradley LA, Palomaki GE, et al., The Evaluation of Genomic Applications in Practice and Prevention (EGAPP) initiative: methods of the EGAPP Working Group, *Genet Med*, 2009;11:3–14.
- Biomarkers Definitions Working Group, Biomarkers and surrogate endpoints: preferred definitions and conceptual framework. *Clin Pharmacol Ther*, 2001;69:89–95.